

## Unexpected Cytological Findings in a Feline Neck Mass

### Contributors

**Sara MEAZZI**<sup>1</sup>, Marianna PANTOLI<sup>1</sup>, Matteo GAMBINI<sup>1</sup>, Antonella RIGILLO<sup>1</sup>, Elisabetta VALSECCHI<sup>2</sup>, Gabriele RATTI<sup>1</sup>

<sup>1</sup>I-Vet Laboratory, Flero, Italy

<sup>2</sup>Animal Care Veterinary Clinic, Medolago, Italy

Sara Meazzi - sara.meazzi@i-vet.it

### Specimen

Fine needle aspiration of a subcutaneous mass located on the right side of the neck, close to the trachea, firm, approximately 1 cm diameter

### Signalment

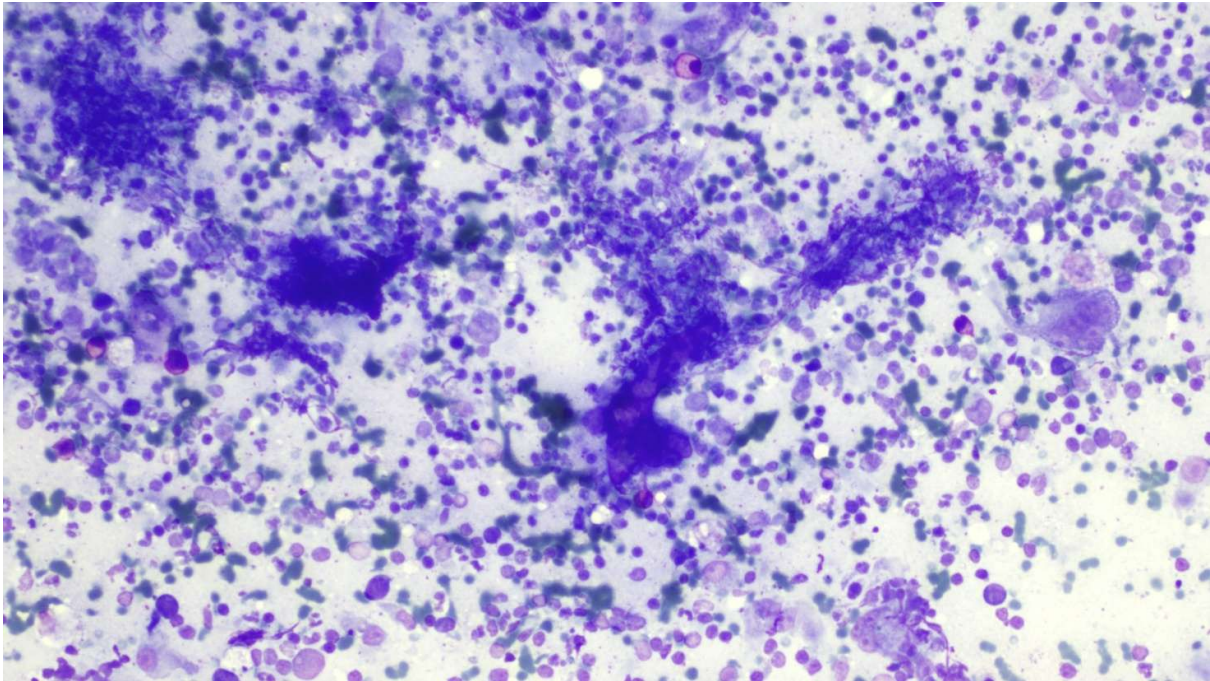
Alessia, Cat, Domestic shortair (DHS), female spayed, 5 years old,

### History

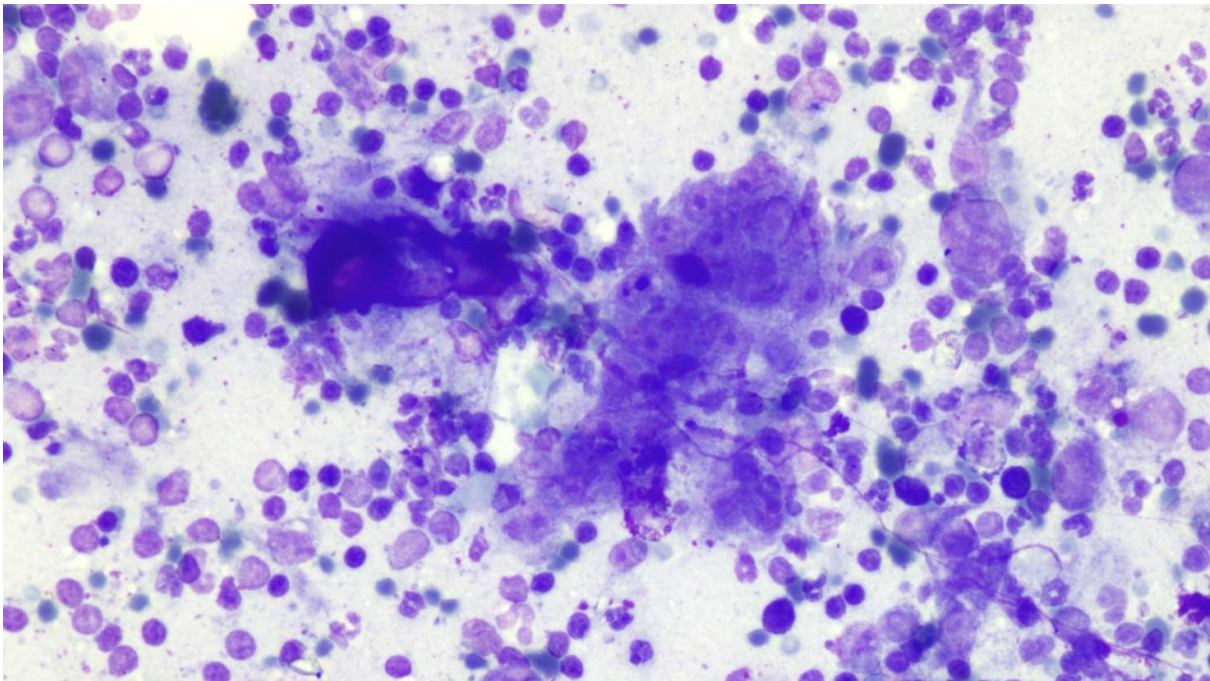
Alessia is an indoor - outdoor cat (access to a private garden), living with another cat and regularly vaccinated. It was asymptomatic except for the presence of a small firm mass located on the right side of the neck, near the trachea.

### **Clinical findings**

Fine needle aspirate (FNA) of the subcutaneous neck mass. Staining: May – Grünwald Giemsa.



*Figure 1. 20x Objective*



*Figure 2. 40x Objective*



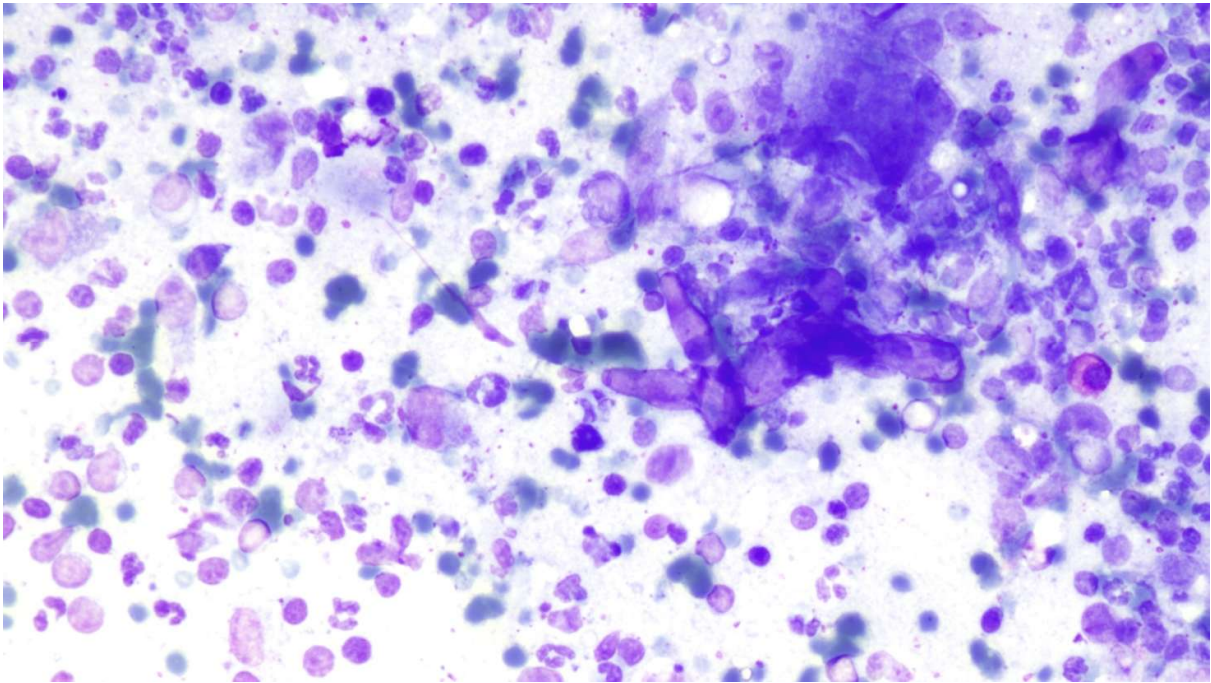


Figure 3. 40x Objective

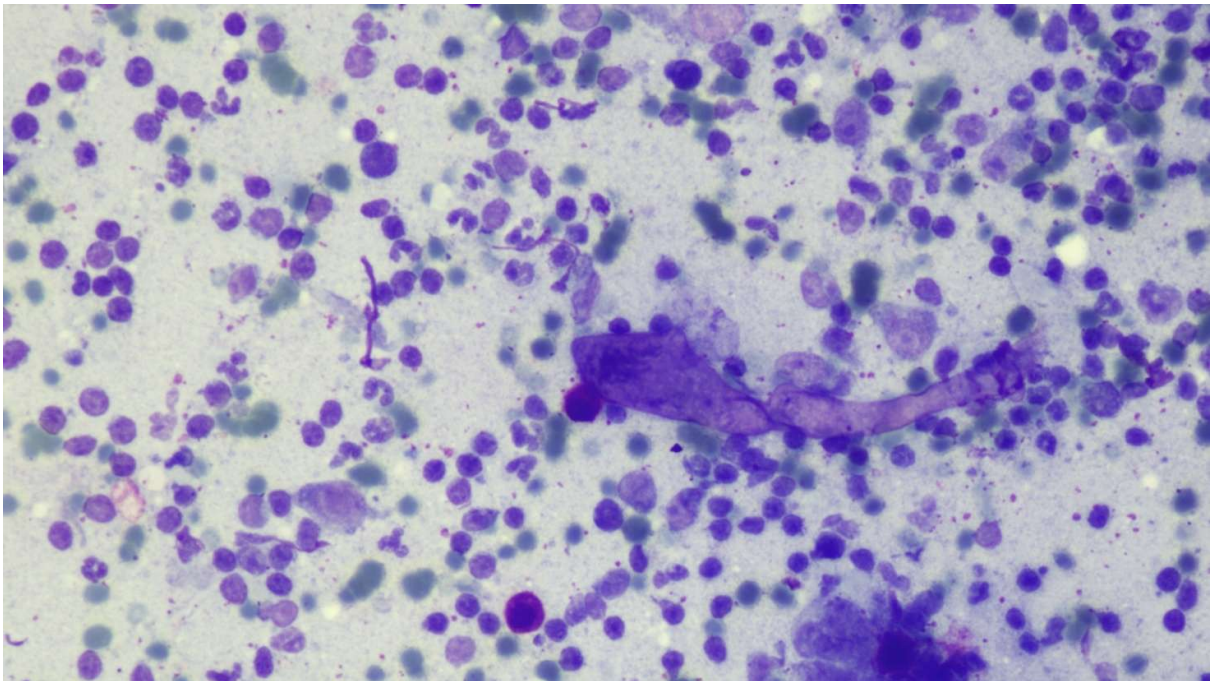


Figure 4. 40x Objective



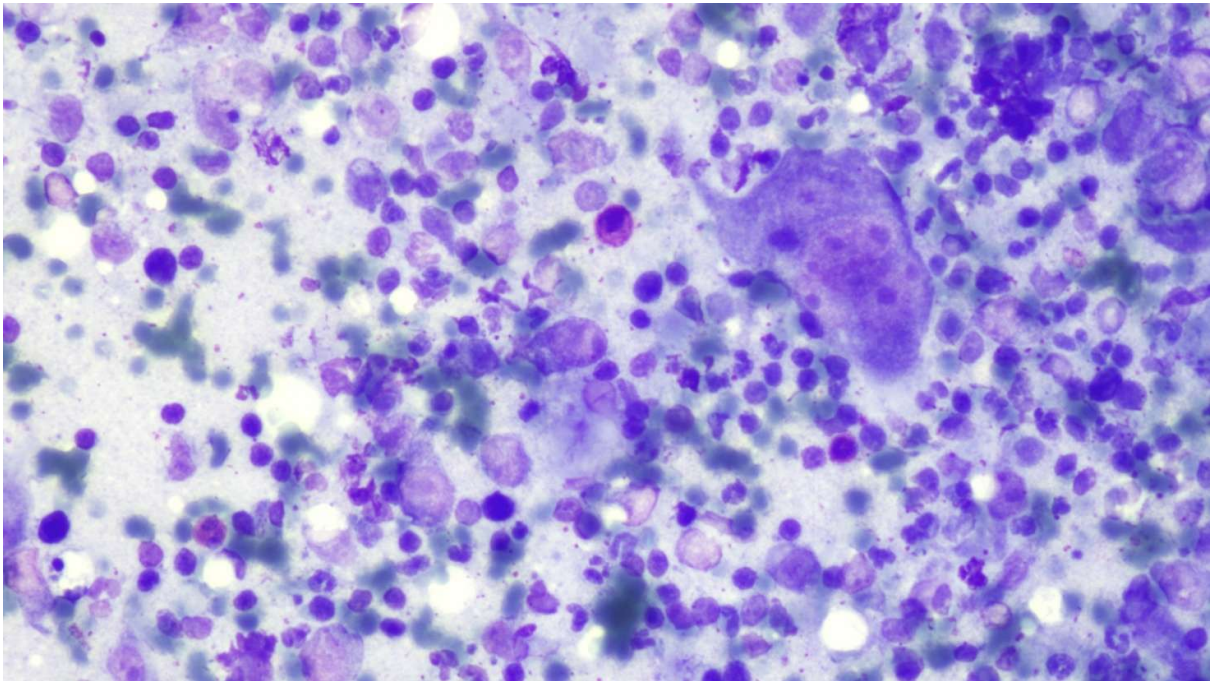


Figure 5. 40x Objective

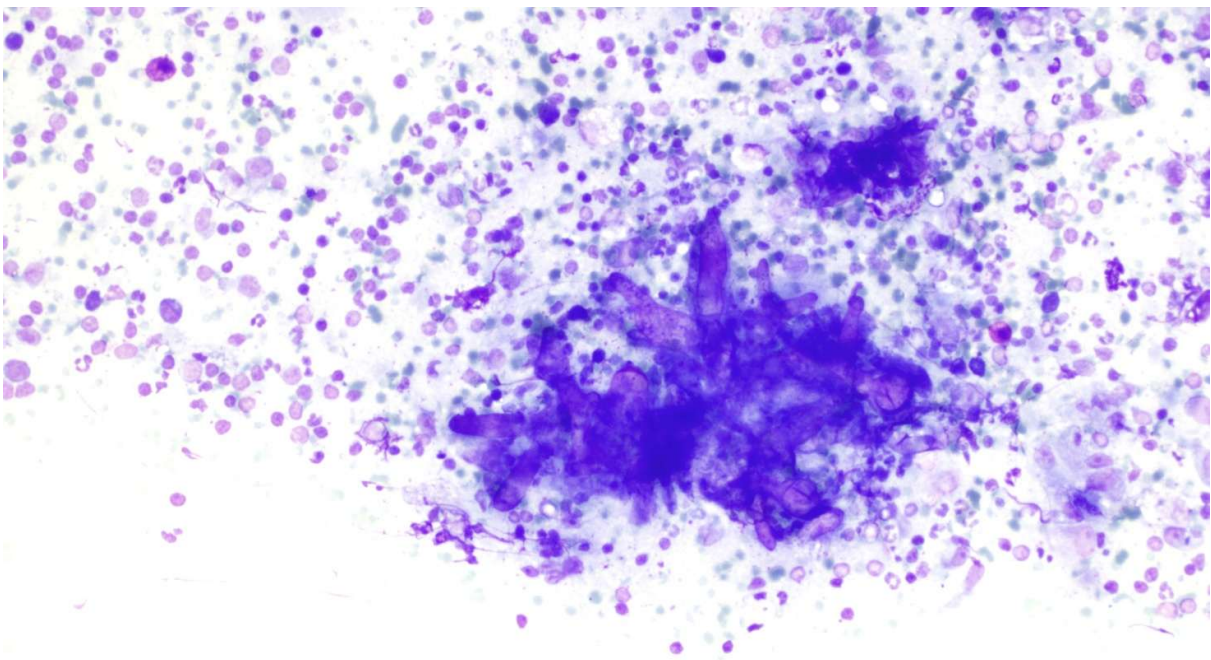


Figure 6. 40x Objective

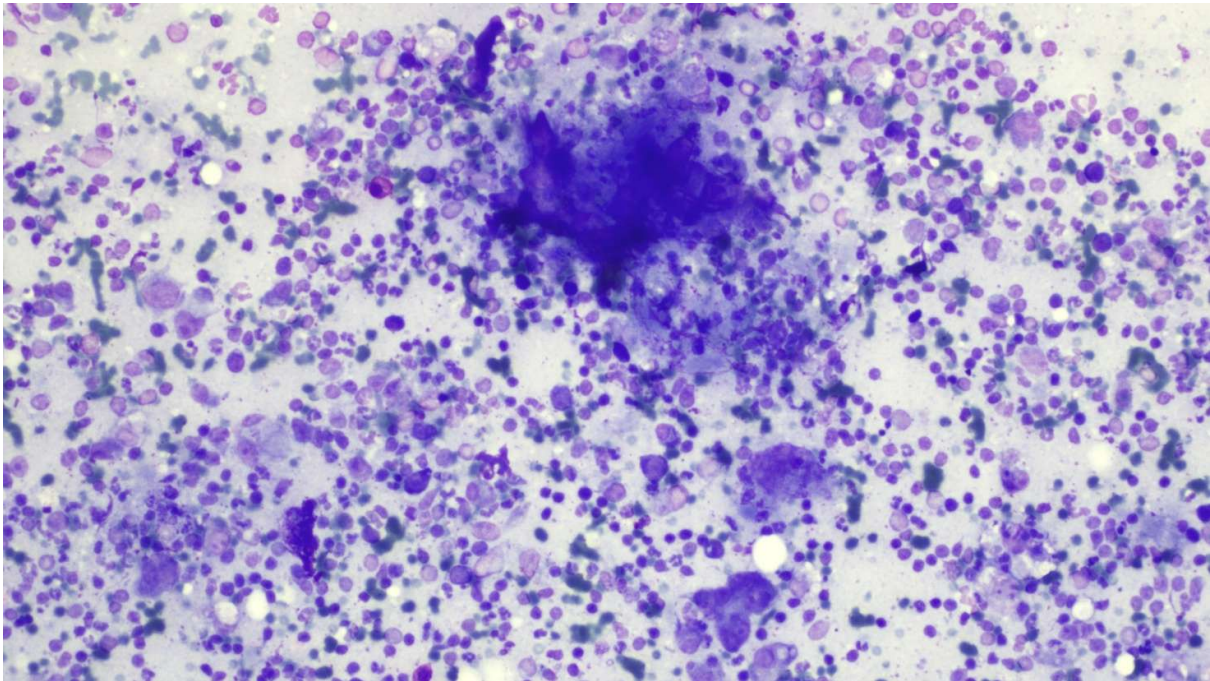


Figure 7. 20x Objective

### Questions

What is your cyological interpretation and diagnosis?

Which further analysis would you recommend?

## **Interpretation/Diagnosis**

The cytological interpretation of the case was the following:

**Description:** The sample had high overall cellularity and good preservation. Moderately bloody background. Presence of a mixed cell population composed mainly by small to medium – sized lymphocytes, followed by a significant proportion of non degenerated neutrophils and macrophages with abundant, slightly basophilic, finely vacuolated cytoplasm. Numerous multinucleated giant cells (up to 30 nuclei) were present, displaying a large amounts of cytoplasm and oval nuclei irregularly arranged. Less numerous were eosinophils, moderately granular mast cells and occasional spindle cells with intensely basophilic cytoplasm and nuclei showing prominent nucleoli. In the sample there were numerous hyphae-like structures, with irregular diameters (5-30 µm), occasionally septate or terminating in globose end. These structures appeared slightly basophilic to amphophilic, sometimes surrounded by a faint capsule that did not take up the stain. They frequently formed large aggregates and were occasionally observed within the cytoplasm of multinucleated cells.

**Cytological diagnosis:** Findings are most consistent with sampling of a lymph node showing pyogranulomatous lymphadenitis, likely secondary to the presence of the aforementioned structures, which are consistent with fungal organisms. Given the lesion's location, a primary fungal granuloma with concurrent sampling of an adjacent lymph node should be considered as a differential diagnosis.

**Comment:** Further histopathological and mycological investigations, along with complete hematological and biochemical profiles, are recommended to assess the lesion's biological behavior and confirm the etiological agent.

## **Answers to questions:**

Cytological findings are consistent with pyogranulomatous lymphadenitis associated with fungal elements. Specifically, histopathology of the neck mass along with mycological examination are suggested to better characterize the etiological agent.



## Additional information

### Histopatological examination

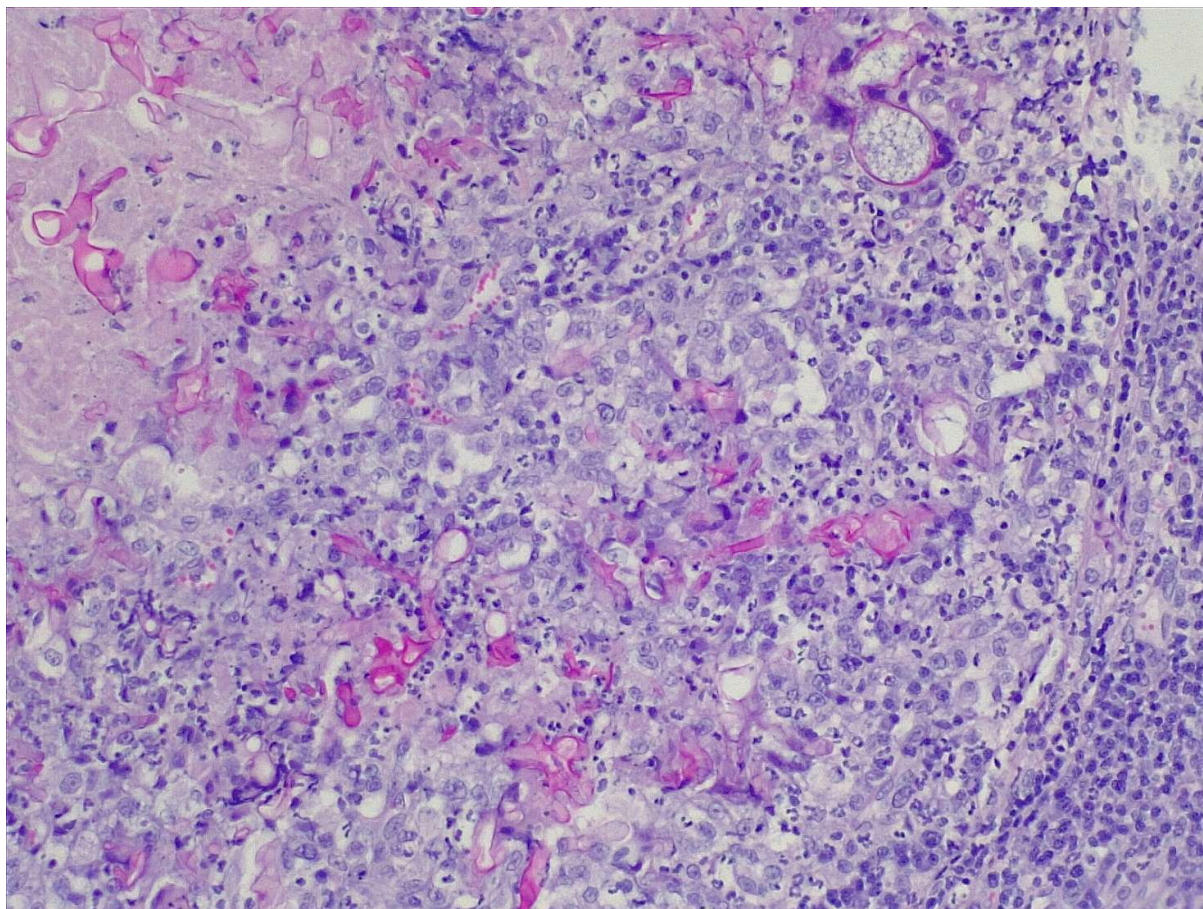


Figure 11. HE staining, 20x Objective.



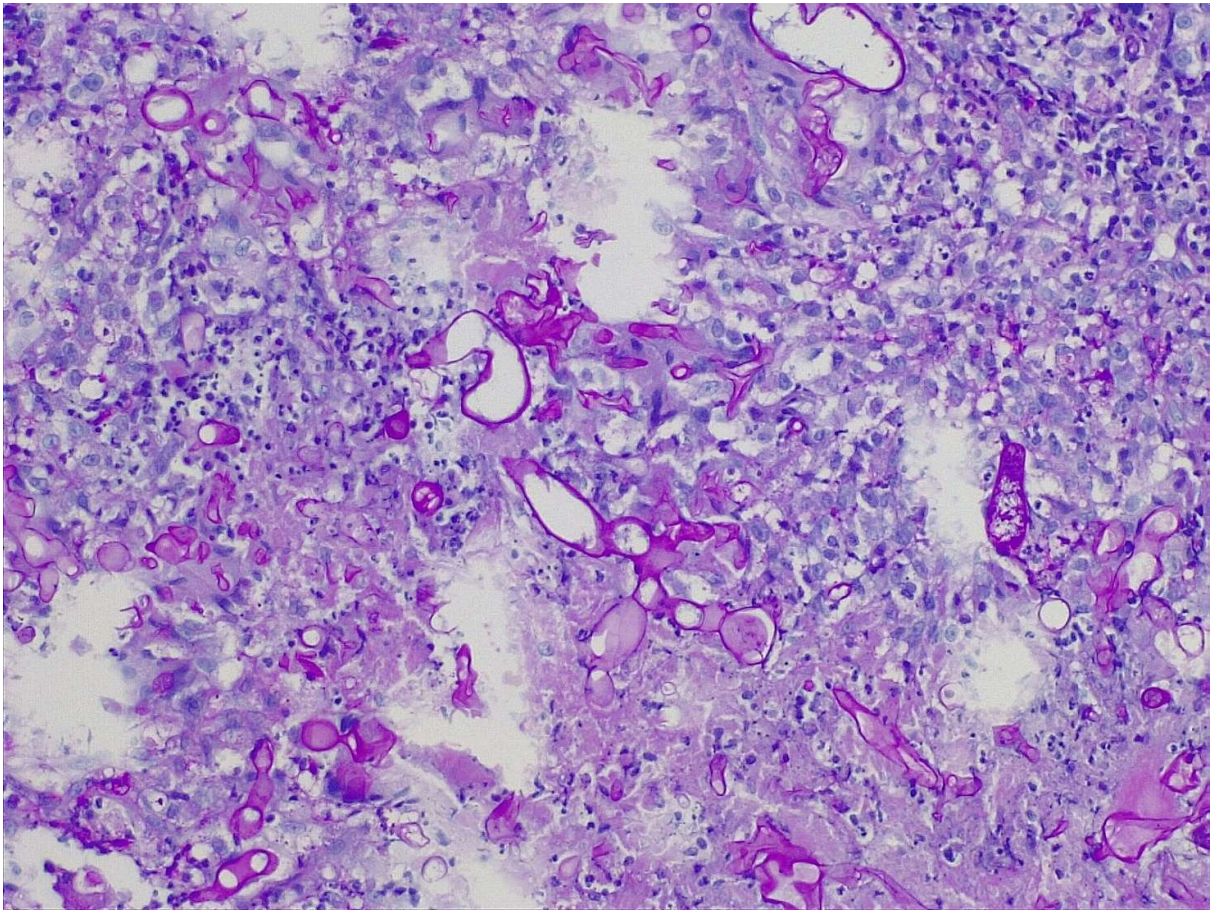


Figure 12. PAS staining, 20x Objective.

Sample: Biopsies of the subcutaneous mass

Macroscopic description: Non-orientable whitish tissue sample, up to 1 cm in diameter.

Microscopic description: Multiple sections of the specimen revealed exclusively coalescing leukocyte aggregates, with central areas composed of necrotic debris admixed with viable and degenerate neutrophils. Numerous fungal structures were observed, consistent with irregularly septate hyphae, characterized by non-parallel walls and non-dichotomous branching, measuring 6–30  $\mu\text{m}$  in thickness and extending over 200  $\mu\text{m}$  in length. Some of them formed "H-shaped" hyphal networks (features consistent with Zygomycetes). The necrosuppurative foci were surrounded by a variably thick rim of epithelioid macrophages, and occasional multinucleated giant histiocytes (pyogranulomas). Peripheral infiltration by small lymphocytes and scattered plasma cells was also noted. PAS staining revealed widespread and variably intense positivity of the fungal hyphae.

Morphological diagnoses: Subcutaneous tissue (based on reported site). Chronic-active pyogranulomatous and lymphoplasmacytic cellulitis with intralesional fungal hyphae consistent with Zygomycetes.

In light of the accompanying mycological findings, the histopathological features are consistent with Mucormycosis (Mauldin, 2016).



## **Mycological analysis**

Sample: Skin biopsy

Result: *Rhizopus* spp.

Comment: *Rhizopus* spp. is a known etiological agent of subcutaneous or nasal mucormycosis. Members of the order *Mucorales* (including *Rhizopus* spp.) are responsible for acute, rapidly progressive infections, particularly in immunocompromised individuals or following traumatic injuries in otherwise healthy individuals (Grooters, 2022).

Fungal colonies grown on culture media were sent to an external laboratory for further fungal typing by sequence analysis. The identified pathogen was *Rhizopus microsporus*.

## **Follow up and clinical outcome**

Two days after cytologic sampling, there was a sudden worsening of the clinical condition. The cat developed purulent nasal discharge, anorexia and an increase in the size of the neck mass. Hematological analysis was performed in house by the referring veterinarian using Idexx ProCyte Dx® (no blood smear examination). Erythrocyte parameters were within reference interval. Concerning the leukocyte, even though the total count was within the reference interval, there was marked neutropenia ( $1.0 \times 10^3/\mu\text{L}$  [2.30 – 10.29]), lymphocytosis ( $12.33 \times 10^3/\mu\text{L}$  [0.92 – 0.67]) and mild monocytosis ( $0.99 \times 10^3/\mu\text{L}$  [0.05 – 0.67]). The analyzer highlighted the possible presence of band neutrophils or immature myeloid forms which required blood smear microscopic confirmation. Mild instrumental thrombocytopenia ( $106 \times 10^3/\mu\text{L}$  [151 – 600]) was also detected. Blood chemistry revealed a moderate hyperproteinemia and hyperglobulinemia (10.5 g/dL [5.7 – 8.9] and 7.7 g/dL [2.8 – 5.1] respectively) without any other abnormalities. FIV and FeLV tests were negative. Radiographs of the neck and thorax showed no significant findings.

One week later, the cat was found comatose and was humanely euthanized by the referring veterinarian.

## **Discussion**

Mucormycosis is an extremely severe fungal disease caused by members of the order *Mucorales*. The most common genera are *Rhizopus* spp. and *Mucor* spp. (Skiada et al., 2017; Czech et Cuellar-Rodriguez, 2025). Among those, *Rhizopus* spp. was the most frequently isolated pathogen and it is mostly associated with an acute and rapidly progressive infection. These are saprophytic molds, ubiquitous in soil or food. In people, mucormycosis typically occurs in immunocompromised patients (e.g. uncontrolled diabetes, hematologic malignancies, transplant patients), though it can also affect immunocompetent individuals following traumatic inoculation (Czech et Cuellar-Rodriguez, 2025). Infection often results from inhalation of sporangiospores that may remain dormant inside macrophages for a long period (Skiada et al., 2017). Common disease manifestations include rhino-orbito-cerebral, pulmonary and cutaneous forms (Czech et Cuellar-Rodriguez, 2025). Cutaneous mucormycosis can be primary (direct skin inoculation) or secondary (dissemination from other locations) (Castrejón-Pérez et al., 2017). Primary lesions may be gradual on onset or fulminant, typically presenting as indurated erythematous to purple plaques that progress to necrosis with

an erythematous halo and eventually eschar formation (Castrejón-Pérez et al., 2017). Secondary forms are more common than the primary ones and generally have a more acute onset with high mortality. The disease generally starts as a sinusitis associated with the presence of necrotic eschar (Castrejón-Pérez et al., 2017). Diagnosis is based on direct microscopy showing broad, nonseptate or pauci – septate, ribbon – like, hyaline hyphae (6 to 25 µm) (Skiada et al., 2017). These fungi tend to be fragile, thus the best mycologic results are obtained by culturing biopsy specimens on regular fungal media incubated at 30 – 37°C (Czech et Cuellar-Rodriguez, 2025). Histopathologic specimens stained with hematoxylin – eosin, periodic acid – Schiff or Grocott – Gomori methenamine silver allow for the identification of hyphae, which are generally associated with necrosis. Molecular identification using MALDI-TOF MS or PCR can confirm the species. (Czech et Cuellar-Rodriguez, 2025)

Direct microscopy is a rapid and cost-effective diagnostic tool that can support early suspicion of fungal infections, particularly in cases with acute clinical progression, demonstrating the presence of fungal elements together with an inflammatory reaction. In this case, cytological examination revealed the presence of fungal elements with features highly suggestive of Mucorales spp. However, direct microscopy may present some limitations. Morphological features may overlap between Mucorales spp. and other filamentous fungi reducing the diagnostic specificity. Moreover, cytologic and histopathological samples may be compromised by poor sample quality, artefacts, or the fragility of the fungal elements themselves (Knoll et al., 2023; Sangoi et al., 2009). Furthermore, they do not allow species-level identification, which may be relevant for therapy (Sangoi et al., 2009). Direct microscopy seems to have an overall accuracy of 79% in the fungal identification (Sangoi et al., 2009). In this case, cytology was essential in triggering further diagnostic efforts, and the definitive identification relied on both histopathology and mycological confirmation, highlighting the importance of a multimodal diagnostic approach.

Cases of mucromycosis are rarely reported in veterinary medicine (Seyedmousavi et al., 2017). In cats, only a few cases are documented. Most feline cases are associated with gastrointestinal involvement (Cunha et al., 2011; Martineau et al., 2023; Mavilio et Bottero, 2025) with two of them linked specifically to *Rizhopus microscoporus*: a gastric case in a young Ragdoll (Mavilio et Bottero, 2025) and a duodenal mass in a 5 year old domestic shorthair cat, the latter diagnosed as feline gastrointestinal eosinophilic sclerosing fibroplasia (FGESF) associated with fungal filaments (Martineau et al., 2023). Another report described *Rhizopus nigricans* in a Ragdoll cat with neurologic signs (Marinelly et al., 2025). All the feline cases showed aggressive clinical courses and high mortality rate, primarily caused by the fungus vasotropic effect, leading to angioinvasion, thrombosis and subsequent tissue necrosis. Only one case of subcutaneous *Mucor* spp. infection was successfully treated with posaconazole for 5 months (Wray et al., 2008). Interestingly, in all the reported feline cases, FIV and FeLV status was negative. This supports the hypothesis that mucormycosis can occur in immunocompetent animals, possibly following unnoticed local trauma, bite wounds, or disruption of cutaneous barriers. The laboratory abnormalities observed in this case included marked neutropenia, lymphocytosis and mild monocytosis, that should be confirmed through blood smear microscopic examination. Neutropenia may be expected in cases of severe acute



inflammatory bacterial or fungal infections. This finding may result from either myeloid exhaustion or peripheral consumption. The suspected presence of immature myeloid forms also suggests a possible left shift, although confirmation via blood smear was not performed. Lymphocytosis may indicate a subacute or chronic inflammatory process in response chronic antigen stimulation. However, this finding also require confirmation on the blood smear, as the automated cell count may have been biased by the presence of immature neutrophils. The presence of increase total protein together with hyperglobulinemia likely reflects a polyclonal immune response to antigenic stimulation. To confirm this hypotesis a serum protein electrophoresis should have been performed. Although these abnormalities are non-specific, they are compatible with the rapidly progressive clinical deterioration observed in this case and are consistent with the immunologic dysregulation frequently associated with invasive mucormycosis. Despite the negative test result for FIV and FeLV, the possibility of transient immunosuppression due to unidentified factors or local trauma cannot be excluded.

## References

- Castrejón-Pérez AD, Welsh EC, Miranda I, Ocampo-Candiani J, Welsh O. Cutaneous mucormycosis. *An Bras Dermatol*. 2017 May-Jun;92(3):304-311. doi: 10.1590/abd1806-4841.20176614.
- Cunha SC, Aguero C, Damico CB, Corgozinho KB, Souza HJ, Pimenta AL, Marassi CD. Duodenal perforation caused by *Rhizomucor* species in a cat. *J Feline Med Surg*. 2011 Mar;13(3):205-7. doi: 10.1016/j.jfms.2011.01.013.
- Czech MM, Cuellar-Rodriguez J. Mucormycosis. *Infect Dis Clin North Am*. 2025 Mar;39(1):121-144. doi: 10.1016/j.idc.2024.11.008
- Grooters AM. Pythiosis, lagenidiosis, paralagenidiosis, entomophthoromycosis, and mucormycosis. In: *Greene's Infectious Diseases of the Dog and Cat* 5<sup>a</sup> ed; 2022; 1105-1117. Elsevier/Saunders.
- Knoll MA, Steixner S, Lass-Flörl C. How to use direct microscopy for diagnosing fungal infections. *Clin Microbiol Infect*. 2023 Aug;29(8):1031-1038. doi: 10.1016/j.cmi.2023.05.012.
- Mauldin EA. Chapter 6: Integumentary System. In: *Jubb, Kennedy & Palmer's pathology of domestic animals: volume 1*; 2016. Elsevier, Inc.
- Marinelly R, Matiassek K, Groth A, Greijdanus-van der Putten S, Rosati M, Tintelnot K, et al. First case of mucormycosis (*Rhizopus nigricans*) in a cat. *Vet Rec Case Rep*. 2025; 13:e1051. <https://doi.org/10.1002/vrc2.1051>
- Martineau M, Tilmant C, Risco Castillo V, Guillot J, Reyes-Gomez E, Benchekroun G, Freiche V. A case of feline gastrointestinal eosinophilic sclerosing fibroplasia associated with fungal

colonisation: endoscopic features, treatment and follow-up. JFMS Open Rep. 2023 May 9;9(1):20551169231165246. doi: 10.1177/20551169231165246.

Mavilio E, Bottero E. Gastric mucormycosis in a cat. JFMS Open Rep. 2025 Feb 12;11(1):20551169241301914. doi: 10.1177/20551169241301914.

Sangoi AR, Rogers WM, Longacre TA, Montoya JG, Baron EJ, Banaei N. Challenges and pitfalls of morphologic identification of fungal infections in histologic and cytologic specimens: a ten-year retrospective review at a single institution. Am J Clin Pathol. 2009 Mar;131(3):364-75. doi: 10.1309/AJCP99OOOZSNISCZ

Seyedmousavi S, Bosco SMG, de Hoog S, Ebel F, Elad D, Gomes RR, Jacobsen ID, Jensen HE, Martel A, Mignon B, Pasmans F, Piecková E, Rodrigues AM, Singh K, Vicente VA, Wibbelt G, Wiederhold NP, Guillot J. Fungal infections in animals: a patchwork of different situations. Med Mycol. 2018 Apr 1;56(suppl\_1):165-187. doi: 10.1093/mmy/myx104. Erratum in: Med Mycol. 2018 Nov 1;56(8):e4. doi: 10.1093/mmy/myy028.

Skiada A, Lass-Floerl C, Klimko N, Ibrahim A, Roilides E, Petrikkos G. Challenges in the diagnosis and treatment of mucormycosis. Med Mycol. 2018 Apr 1;56(suppl\_1):93-101. doi: 10.1093/mmy/myx101.

Wray JD, Sparkes AH, Johnson EM. Infection of the subcutis of the nose in a cat caused by *Mucor* species: successful treatment using posaconazole. J Feline Med Surg. 2008 Oct;10(5):523-7. doi: 10.1016/j.jfms.2008.06.001.